Review

Antitumor Activity of Platinum Complexes

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Summary. Contemporary ideas about the mechanism of antitumor activity of platinum complexes are reviewed and discussed. The induction of SOS functions in bacteria is emphasized and an analogous mechanism in animal cells is suggested. The fate of the leaving ligand in the body is not known. Therefore the complex which reaches the target DNA may be very different from the applied compound. Even the amino ligand may be detached in the body, probably as part of the detoxication by binding to proteins. In the case of fast or medium-speed reactive complexes most of the platinum is inactivated by binding to proteins, whereas slow-reacting drugs are mostly excreted unchanged in the urine. Thus, the quantity of the complex which reaches the target ist also difficult to assess. Due to peculiar pharmacokinetics, the results obtained with poorly soluble compounds administered in the solid form cannot be compared with those obtained in true solutions. There are many reasons for believing that the study of a coordination anticancer drug may contribute to our understanding of cancer growth and its reversal.

Introduction

Platinum complexes represent quite an unusual group of drugs; cisplatin [cis-diamminedichloroplatinum(II)] is the first inorganic antitumor drug to be used in clinics. It belongs to a class of very simple molecules, being composed of eleven atoms, six of them hydrogen. Cisplatin is important not only because of its good clinical record, but also because it is the parent compound of a new class of drugs. The study of mechanism of action of the metal coordination compounds may provide new insight into the nature of malignant growth and its reversal. It is the aim of this review to summarize facts and theories from this point of view.

The Primary Reaction

The chemical similarity of cisplatin to nitrogen mustard-type alkylating agents led to the hypothesis [10, 27, 29] that direct attack on DNA, namely interstrand cross-linking, might be the primary reaction responsible for the specific biological effect. Detailed studies [4, 13, 39, 45, 47, 70, 85] have shown that

a) the cytostatic effect is roughly proportional to the number of Pt atoms bound to the DNA [64], and to the extent of cross-linking [54, 55, 86];

- b) the repair system may partially abolish the effect, probably by the well-known excision mechanism [5, 17, 68, 69];
- c) these rules hold only for the cis isomer.

The difference between cis and trans isomers is of key importance for understanding the mechanism of action, and many studies have been devoted to this problem [21, 33, 45, 47, 48, 54, 64, 68, 70, 75, 79, 84–86]. It is obvious that the strict requirement for cis configuration cannot be explained by the reactivity of the isomers, since in some systems the trans isomer binds more effectively to the DNA than the cis isomer [2]. The reaction of the first ligand of the cis or trans isomer is almost identical, whereas the reaction of the other ligand, which follows after some delay [84], brings the specifity. The trans isomer forms more DNA-protein cross-links, while the cis isomer forms interstrand bridges within the DNA [33] and also causes much faster unwinding [74]. However, the critical alteration of the structure of DNA which is responsible for the absolute specifity of the cis position of the leaving ligands still remains unidentified [2, 7, 19, 74]. Model studies show there is a wide variety of possible products. It may be that there are many modes of attack on DNA [18, 31, 47]. Nevertheless, the hypothesis about intercalation of the planar complexes similar to ethidium seems to be ruled out [71].

Emergency State

As for many other cytostatic reagents (viz., alkylating agents and radiation) inhibition of DNA snythesis [23, 28], restraint of nuclear [24, 57] and cellular division [6, 76], and occurrence of mutations and chromosomal abberations [44, 52, 83] are the general phenomenological responses to the primary reaction. In bacteria it has been well established that cisplatin-induced filamentous growth [72], as well as the well-known induction of bacteriophages [50, 66] and mutagenesis [12] are the results of the so-called SOS functions [12, 32, 49, 73]. SOS functions is a name for a sequence of emergency reactions [63] programmed by several genes and triggered by the occurrence of a single-stranded region in the DNA (ss-DNA) [14] which results from the replication of damaged DNA matrix. The ss-DNA activates the enzymogen called recA protein (the name is derived from its involvement in DNA recombination), which gains specific proteolytic affinity for so-called lexA protein. The latter acts as a repressor of several genes controlling cell division and DNA replication and repair, and also genes coding for recA and lexA proteins. In this regard, SOS functions represent an alarm system with built-in amplification

by positive feedback. Several prophages join this system by shaping the repressor of their induction so close to the structure of the lexA protein that it is also split by the activated recA protease. An extensive review is available on this aspect [14].

No such detailed description of emergency processes triggered in an animal cell by specific DNA damage is yet available. There are, however, enough data supporting the idea that in animal cells also the specific damage to the DNA evokes processes having biological role similar to the SOS functions. For example, the treatments (irradiation, chemicals) which exhibit cytostatic (antitumor), mutagenic or cancerogenic effects in animal models induce SOS functions in bacteria. In fact, the induction of prophages was suggested as a screening method for antitumor drugs [41, 62] and carcinogens [26]. Specific proteolytic activity [34, 46], protein phosphorylation [67], and carboxymethylation [53], which is parallel in its effect to repressor inactivation in bacteria, were shown to play a role in some of the processes caused by treatment with platinum complexes: viz. the induction of high cAMP level [22], the expression of oncogenic virus markers [82], transformation [52], mutation [44], gene [40] and cell-surface rearrangement [59], growth cessation [38], etc. Thus, we may expect that in the near future an emergency state analogous to SOS functions will be described in animal cells, which will account for specific response to the treatment by cisplatin and analogs. The difference in DNA damage by cis and trans isomers when properly studied may provide a valuable tool.

The Role of the Leaving Ligand

It is well known that the two chlorine atoms in cisplatin can be easily replaced, even by water [42, 65]. The suggested reaction with DNA consists in their replacement by the nitrogen and, maybe, oxygen of purine bases [33, 45, 47, 48, 71, 80]. Thus, the Pt(II) complex may be active only when the *cis*-oriented leaving ligands Pt-X are thermodynamically less stable than the Pt-N ligand [81]. Therefore, the replacement of the ligand by nitrogen or sulfur (which are the platinum binding atoms, e.g., in DNA and proteins) results in effectively irreversible inactivation [25, 57].

In the collection of cisplatin analogs called second-generation drugs we may find leaving ligands of different stability and reactivity: very reactive, e.g., sulfate, or quite stable, e.g., chelating malonate. We may expect that the former ligands are very quickly replaced after the introduction in the body by more abundant species, e.g., Cl-, OH-, H₂O, giving rise to other active complexes. However, the reactive ligands also increase the probability of the above-mentioned irreversible inactivation by reacting with nitrogen or sulfur atoms in various components of the biological milieu. The chelating dicarboxylates are often considered [10] as very stable; and corresponding complexes are called 'prodrugs' and are active only upon enzymatic transformation [11]. We were able to show [16] that at physiological pH and ionic strength malonato derivatives react, probably after the dissociation of the carboxylate, but only slowly. Since the biological response to them is as fast as with cisplatin, the problem of chelating leaving ligands cannot be considered clear.

Replacement of the ligand after introduction of the drug into the body has the following consequences:

- a) we do not know which leaving ligand is present in the complex when it reaches the target cancer cell;
- b) we do not know which ligand is attached to the platinum atom when it interacts with DNA inside the target cell;c) the reactivity of the leaving ligand probably determines the fraction of the drug which is inactivated before it reaches the

Originally we suggested that the complex which penetrates the cell must be neutral [15]. Now there are some second-generation drugs which at physiological pH carry either a positive (e.g., monoglucuronato complex) [36], or a negative (e.g., carboxyphtalato complex) [20] charge. Nevertheless, the theory of neutrality may still be valid, since the ligand responsible for the charge may be replaced before the complex reaches the target cell.

Thus, the nature of the leaving ligand in the administered drug controls the fraction of the active platinum complex which reaches the target, but it is not certain whether it plays any role in penetration into the target cell and in the reaction with the DNA in it. On the other hand, determining the nonspecific interactions it is responsible for many side-effects and toxicities. On the basis of available data we can suggest that slowly reacting drugs are free of nephrotoxic side-effects, since they pass the kidney without reacting with functional structures (e.g., -SH groups) in it. This is reflected by quite a large fraction of unchanged drug in the urine (the same is true for Pt(IV) drugs; see below).

The Fate of the Amino Ligand

In contrast to the situation with the anionic ligand, a charge on the amino ligand abolishes the activity of the complex [30]. In fact it seems [11] that the more hydrophobic the amine the more active is the complex which results. There should be at least one hydrogen atom attached to the nitrogen, i.e., tertiary amines do not provide active complexes, which suggests an involvement of hydrogen bridges in the primary reaction [56]. Since there is less freedom in the choice of physical and chemical properties of the amino ligand we may conclude that it accompanies the platinum atom to the target DNA, being in this way the main factor determining the pharmacokinetics. It may also affect penetration into the cell and interaction with the DNA.

This is not to say that detachment of the amino ligand may not occur in vivo [77; A. B. Robins and A. O. Leach, unpublished work]. According to the chemical rules it may result from the *trans* effect after replacement of the leaving ligand by sulfur. In other words, the detachment of the amino ligand may be a part of the inactivation reaction and thus it would be irrelevant to the primary reaction. However, from the practical point of view it would make it impossible to use labeled amines as tracers in pharmacokinetic studies of the second-generation drugs [29, 51].

Another source of serious mistakes might be the tendency of bulky amino ligands to undergo *cis-trans* isomerization under just recrystallization [8].

The Solubility Problem

As mentioned above, it seems that lipophilic amines, particularly those with a branched or cycloalkyl hydrocarbon part,

provide better drugs than more hydrophilic amines [10]. Naturally, the water-solubility of complexes decreases with increasing hydrophobicity of the amine, and valid data on toxicity and activity can be obtained only with drugs given in the form of true solution. Any 'slurry' or 'sonicates' with supporting polymers (carboxymethyl cellulose, polyvinylpyrrolidone, polyvinalcohol, etc.) are misleading. They represent a drug depot which slowly releases the soluble drug – the only form which can reach and penetrate the target cell. The rate of release is unpredictable due to unknown particle size, effects of additives, etc. After IP administration such microparticles are subjected to phagocytosis by the macrophages, which complicates the pharmacokinetics even further. Particulary when the model tumor is in the ascitic form, which is also administered IP, the results are highly artificial. Even with true solution, IP administration (sometimes necessary because of large volumes) may result in some artefacts [61] in the drug deposition.

The use of DMSO as the solvent can bring about only a limited solution of this problem, since it replaces the leaving ligand, thus inactivating the drug [35].

Drugs Based on Tetravalent Platinum

The Pt(II) complexes can be oxidized giving rise to the octahedral structures with two additional ligands above and below the plane of the original Pt(II) complex. These additional ligands are preferentially hydroxyls, since they increase the solubility, whereas others, e.g., chlorines, make the complex less soluble.

It follows from the electronic d⁶ structure that the Pt(IV) complexes are very stable. The equatorial ligands cannot be replaced under physiological conditions and the out-of-plane ligands only very slowly, if at all [9, 78]. Thus, such complexes cannot exhibit effect until after returning to the original Pt(II) form, i.e., after reduction.

Early predictions said [81] that reduction in vivo must be quite a fast process, so that Pt(IV) drugs were more soluble forms of the Pt(II) analogs and that the pharmacokinetics of Pt(IV) need not be considered. However, when the optimal dose for the Pt(IV) derivatives is compared with the toxic dose of the parent Pt(II) complex it becomes evident that the former is usually several times higher than the latter [60]. This indicates that reduction is by no means an instantaneous event and that a substantial part of the dose of the Pt(IV) complex must be eliminated or detoxicated.

Indeed, new data show [58] that reduction in the body does not proceed very quickly, so that great fraction of the IV-administered Pt(IV) complex is excreted in the urine unchanged. Thus, the Pt(IV) complex may be regarded as an inert depot form of an active Pt(II) reduction product. Such very convenient substitution of Pt(IV) drug for the long-term infusion will be compensated by some loss of the complex in urine. In this regard Pt(IV) complexes seem to be very attractive drug forms.

However, the first data obtained with CHIP, cis-dichloro-bis(isopropylamine)-trans-dihydroxy platinum(IV), indicated [58] one problem: even in well-selected experimental beagle dogs wide individual differences were observed in the reduction rate. If future studies confirm this observation it may represent a serious drawback to the clinical use of Pt(IV) drugs, since with human patients even more pronounced individual differences in reduction rate can be expected, i.e., in the formation of the active and also toxic form of the drug. Thus, biochemistry and localization of the reduction reaction deserve careful consideration.

Other Biological Effects of the Platinum Complexes

In this review we have discussed only the antitumor activity of platinum complexes. Even here other theories have been suggested. Aggarwal et al. [1] found damage to mitochondria, oxidative phosphorylation, altered calcium balance and microfilament organization. Hall et al. [22] observed an increase of cAMP and a corresponding decrease of histone phosphorylation. There are many other observations of metabolic effects of platinum drugs [3, 22].

In general, we may discriminate three classes of effects:

- 1) The cytostatic effect described above, which is close to the action of radiation and of alkylating substances and is caused by very specific damage to the DNA. It represents the basis for the antitumor and mutagenic activity, gastrointestinal toxicity, and myelotoxicity;
- 2) Metabolic effects, which follow the general pattern of heavy metal poisoning, probably caused by the reaction with -SH groups in some important enzymes (e.g., ATPase) [3]. This may be the basis of the nephrotoxicity [43];
- 3) A poorly documented and little understood membrane effect, which may be observed at concentrations much lower than the above effects (of the order 10⁻⁹ molar). It is responsible for, e.g., the stimulation of spontaneous monocyte-mediated cytotoxicity [37].

The different nature of the antitumor and nephrotoxic activity give rise to hope that active analogs may be found that are free of nephrotoxic side-effects. Unfortunately, the same cannot be said of myelotoxicity and gastrointestinal injury.

It has not been possible in this review to discuss all facts known about platinum antitumor drugs. Therefore we have chosen only those we consider most important for further developments in this field.

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References

- Aggarwal SK, Whitehouse MW, Ramachandran C (1980) Ultrastructural effects of cisplatin. In: Cisplatin, current status and new developments. Academic Press, New York, p 79
- Alazard R, Germanier M, Johnson NP (1982) Mechanism of toxicity of platinum(II) compounds in repair-deficient strains of Escherichia coli. Mutat Res 93: 327
- Aull JL, Allen RL, Bapat AR, Daron HH, Friedman ME, Wilson JF (1979) The effects of platinum complexes on seven enzymes. Biochim Biophys Acta 571: 352
- Barton JK, Lippard SJ (1980) Heavy metal interactions with nucleic acids In: Nucleic acid – metal ion interactions. John Wiley, New York, p 31
- Beck DJ, Brubaker RR (1973) Effect of cis-platinum(II)-diamminedichloride on wild-type and deoxyribonucleic acid repair-deficient mutants of Escherichia coli. J Bacteriol 116: 1245
- Bergerat J-P, Barlogie B, Gohde W, Johnston DA, Drewinko B (1979) In vitro cytokinetic response of human colon cancer cells to cis-dichlorodiammine-platinum(II). Cancer Res 39: 4356

- Botour JL, Macquet JP (1981) Viscosity, nicking, thermal and alkaline denaturation studies on three classes of DNA-platinum complexes. Biochim Biophys Acta 653: 305
- 8. Bradford JP, Faggini R, Lock CJL (1981) trans-Dichlorobis(cyclopentylamine)platinum II. Acta Crystalogr [B] 37:243
- Budanova NS, Zheligovskaya NN, Spicyn VI (1978) The synthesis and the study of some physico-chemical properties of acidocomplexes of platinum(IV). Koordin Chim 4:930
- Cleare MJ, Hoeschele JD (1973) Antitumor platinum compounds

 relationship between structure and activity. Platinum Metal Rev
 17:187
- Cleare MJ, Hydes PC, Hepburn DR, Malerbi BW (1980) Antitumor platinum complexes: Structure-activity relationship.
 In: Cisplatin, current status and new developments. Academic Press, New York, p 149
- 12. Cunningham D, Pembroke JT, Stevens E (1981) cis-Platinum(II) diamminodichloride-induced mutagenesis in E. coli K12: crowding depression of mutagenesis. Mutat Res 84: 273
- 13. Deutsch WA, Spiering AL, Newkome GR (1980) An in vitro characterization of interstrand cross-links in DNA exposed to the antitumor drug *cis*-dichlorodiammineplatinum(II). Biochem Biophys Res Commun 97:1220
- Devoret R (1981) Inducible error-prone repair and induction of prophage lambda in *Escherichia coli*. Prog Nucleic Acid Res Mol Biol 26: 251
- Drobnik J, Horáček P (1973) Specific biological activity of platinum complexes. Contribution to the theory of molecular mechanism. Chem Biol Interact 7: 223
- Drobník J, Nosková D (1982) The reactivity of malonato platinum(II) complexes. XXII International Conference of Coordination Chemistry, Abstract ThP 74
- 17. Drobník J, Urbánková M, Krekulová A (1973) The effect of cis-dichlorodiammineplatinum(II) on Escherichia coli B. The role of fil, exr and hcr markers. Mutat Res 17:13
- 18. Eastman A (1982) Comparison of the interaction of *cis* and *trans*-diamminedichloroplatinum(II) with DNA by a simple filter binding assay. Biochem Biophys Res Commun 105: 869
- Erickson LC, Zwelling LA, Ducore JM, Sharkey NA, Kohn KW (1981) Differential cytotoxicity and DNA cross-linking in normal and transformed human fibroblasts treated with cis-diammine-dichloroplatinum(II). Cancer Res 41:2791
- Gale GR, Smith AB, Schwartz P (1979) Preliminary studies of 4-carboxyphtalato(1.2-diaminocyclohexane) platinum(II): antitumor activity and effects on macromolecular synthesis. J Clin Hematol Oncol 9:217
- 21. Ganguli PK, Theophanides T (1981) Premelting phenomenon in DNA caused by the anti-tumor drug *cis*-dichlorodiamminoplatinum. Inorg Chim Acta 55: L43
- Hall IH, Holshouser MH, Loeffler LJ (1980) Effects of cis-malonato-diammino platinum(II) on P-388 lymphocytic leukemia cell metabolism. J Pharm Sci 69: 1160
- 23. Harder HC, Rosenberg B (1970) Inhibitory effects of anti-tumor platinum compounds on DNA, RNA and protein synthesis in mammalian cells in vitro. Int J Cancer 6:207
- Heinen E, Bassleer R (1976) Mode of action of cis-dichloro-diammine platinum(II) on mouse Ehrlich ascites tumour cells. Biochem Pharmacol 25: 1871
- 25. Heinen E, Bassleer R, Desaive C (1978) Effects of antimitotic agents bound to a macromolecular carrier in normal or cancer cells. Importance of the mode of binding of the agent to the carrier. Eur J Cancer 14:1005
- Ho YL, Ho SK (1981) Screening of carcinogens with the prophage λcIts857 induction test. Cancer Res 41:532
- 27. Horáček P, Drobník J (1971) Interaction of *cis*-dichlorodiammine platinum (II) with DNA. Biochim Biophys Acta 254: 341
- 28. Howle JA, Gale GR (1970) cis-Dichlorodiammineplatinum(II): persistent and selective inhibition of deoxyribonucleic acid synthesis in vivo. Biochem Pharmacol 19: 2757
- Howle JA, Gale GR, Smith AB (1972) A proposed mode of action of antitumor platinum compounds based upon studies with

- cis-dichloro(dipyridine)platinum(II). Biochem Pharmacol 21: 1465
- Inagaki K, Kidani Y, Suzuki K, Tashiro T (1980) Platinum complexes of diaminocarboxylates and their ethyl ester derivatives: antitumor activity and interaction with deoxyribonucleic acid. Chem Pharm Bull 28: 2286
- Johnson NP (1982) Preliminary characterization of the adducts formed between the antitumor compound cis-Pt(NH₃)₂ Cl₂ and DNA. Biochem Biophys Res Commun 104: 1394
- Kelehan M, Farrel E, Smith P (1977) Induction of SOS functions by cis-platinum diamminodichloride. Proc Soc Gen Microbiol 4: 135
- Kelman AD, Peresie HJ, Stone PJ (1977) An analysis of the modes of binding of antitumor platinum complexes to DNA. J Clin Hematol Oncol 7: 440
- 34. Kennedy AR, Little JB (1981) Effects of protease inhibitors on radiation transformation in vitro. Cancer Res 41:2103
- 35. Kerrison JS, Sadler PJ (1977) Solvolysis of cis Pt(NH₃)₂Cl₂ in dimethyl sulphoxide and reaction of glycine with PtCl₃(Me₂SO) as probed by ¹⁹⁵Pt nuclear magnetic resonance shifts and ¹⁹⁵Pt ¹⁵N coupling constants. J Chem Soc/Chem Commun 1977: 861
- Kidani Y, Noji M, Tsakagoshi S, Tashiro T (1978) Antitumor activity of water-soluble platinum(II) complexes of 1,2-cyclohexanediamine isomers. Gan 69: 263
- Kleinerman ES, Zwelling LA, Muchmore AV (1980) Enhancement of naturally occurring human spontaneous monocyte-mediated cytotoxicity by cis-diamminedichloroplatinum(II). Cancer Res 40: 3099
- Kletzien RF, Miller MR, Pardee AB (1977) Unique cytoplasmatic phosphoproteins are associated with cell growth arrest. Nature 270: 57
- Laurent G, Erickson LC, Sharke NA, Kohn KW (1981) DNA cross-linking and cytotoxicity induced by cis-diamminodichloroplatinum(II) in human normal and tumor cell lines. Cancer Res 41: 3347
- Lavi S (1981) Carcinogen-mediated amplification of viral DNA sequences in simian virus 40-transformed Chinese hamster embryo cells. Proc Natl Acad Sci USA 78: 6144
- Lein J, Haidemann B, Courevitch A (1962) Induction of lysogenic bacteria as a method of detecting potential antitumor agents. Nature 196: 783
- 42. LeRoy AF (1979) Some quantitative data on *cis*-dichlorodiam-mineplatinum(II) species in solution. Cancer Treat Rep 63:231
- 43. Levi J, Jacobs C, Kalman SM, McTigue M, Weiner MW (1980) Mechanism of *cis*-platinum nephrotoxicity: I. Effects of sulfhydryl groups in rat kidneys. J Pharm Exp Ther 213: 545
- 44. Levine BS, Preache MM, Pergament E (1980) Mutagenic potential of *cis*-dichlorodiammine platinum in rodents. Toxicology 17:57
- 45. Lippard SJ (1980) Binding of platinum antitumor drug to its likely biological targets. In: Inorganic chemistry in biology and medicine, p 147 (ACS symposium series no 140)
- Long CW, Bruszewski LA, Snead RM (1980) Analysis of transcription during type C viral induction using cell permeabilization techniques. Cancer Res 40: 22
- Macquet JP, Botour JL (1978) Modifications of the DNA secondary structure upon platinum binding: a proposed model. Biochimie 60: 901
- 48. Macquet JP, Theophanides T (1975) DNA-platinum interactions in vitro with *trans* and *cis*-Pt(NH₃)₂Cl₂. Bioinorgan Chem 5:59
- 49. Markham BE, Brubaker RR (1980) Influence of chromosome integrity on *Escherichia coli* cell division. J Bacteriol 143: 455
- Monti-Bragadin C, Ramani L, Samer L, Mestroni G, Zassinovich G (1975) Effects of cis-dichlorodiammineplatinum(II) and related transition metal complexes on Escherichia coli. Antimicrob Agents Chemother 7: 825
- 51. Morris CR, Gale GR (1973) Interaction of antitumor platinum compound with deoxyribonucleic acid, histones, L-amino acids,

- poly-1-amino acids, nucleosides and nucleotides. Chem Biol Interact 7:305
- 52. Morrison WD, Huff V, Colyer SO, DuFrain RJ, Littlefield LG (1981) Cytogenetic effects of *cis*-platinum(II)diamminedichloride in vivo. Environmental Mental Mutagenesis 3:265
- O'Dea RF, Vineros OH, Diliberto EG (1981) Protein carboxymethylation: role in the regulation of cell functions. Biochem Pharmacol 30: 1163
- 54. Pascoe JM, Roberts JJ (1974a) Interactions between mammalian cell DNA and inorganic platinum compounds. I. DNA interastrand cross-linking and cytotoxic properties of platinum(II) compounds. Biochem Pharmacol 23:1345
- Pascoe JM, Roberts JJ (1974b) Interactions between mammalian cell DNA and inorganic platinum compounds. II. Interstrand cross-linking of isolated and cellular DNA by platinum(IV) compounds. Biochem Pharmacol 23:1359
- 56. Pasini A (1982) A stereochemical investigation on the deprotonation of guanosine coordinated to the cis-Pt moiety. XXII International Conference on Coordination Chemistry. Abstract ThP42
- 57. Pauw-Gillet MC, Houssier C, Fredericq E (1979) Interaction of DNA and purine nucleoside with *cis*-dichlorodiammineplatinum(II) and antimitotic activity of the complexes on meristematic root cells. Chem Biol Interact 25:87
- 58. Pendyala L, Cowens JW, Creaven PJ (1982) Studies on the pharmacokinetics and metabolism of *cis*-dichloro*trans*dihydroxy-bis-isopropylamine platinum(IV) in the dog. Cancer Treat Rep 66: 509
- Prasad SB, Sodhi A (1981) Effect of cis-dichlorodiammine platinum(II) on the agglutinability of tumor and normal cells with concavaline A and wheat germ agglutinin. Chem Biol Interact 36:355
- Prestayko AW, Bradner WT, Huftalen JB, Rose WC, Schuring JE, Cleare MJ, Hydes PC, Crooke ST (1979) Antileukemic (L1210) activity and toxicity of cis-dichlorodiammineplatinum(II) analogs. Cancer Treat Rep 63: 1503
- Oretorius RG, Petrilli ES, Kean C, Ford LC, Hoeschele JD, Lagasse LD (1981) Comparison of the iv and ip routes of administration of cisplatin in dogs. Cancer Treat Rep 65: 1055
- 62. Price KE, Buck RE, Lein J (1964) System for detecting inducers of lysogenic *Escherichia coli* W 1709 (λ) and its applicability as a screen for antineoplastic antibiotics. Appl Microbiol 12: 428
- 63. Radman M (1975) SOS repair hypothesis: phenomenology of an inducible DNA repair which is accompanied by mutagenesis In: Molecular mechanisms for repair of DNA, part A. Plenum, New York, p 355
- 64. Rahn RO, Johnson NP, Hsie AW, Lemontt JF, Masker WE, Regan JD, Dunn WC, Hoeschele JD (1980) The interaction of platinum compounds with the genome: Correlation between DNA binding and biological effects. In: The scientific basis of toxicity assessment. Elsevier/North Holland Biomedical Press, Amsterdam, p 153
- 65. Reishus JN, Martin DS (1961) cis-Dichlorodiammine platinum(II): Acid hydrolysis and isotopic exchange of the chloride ligands. J Am Chem Soc 83: 2457
- 66. Reslová S (1972) The induction of lysogenic strains of Escherichia coli by cis-dichlorodiammineplatinum(II). Chem Biol Interact 4:66
- 67. Rieber MS, Rieber M (1981) Transformed cells exhibit altered response to DB cyclic AMP-mediated modulation of protein phosphorylation and different endogenous phosphoprotein acceptors. Cancer Biochem Biophys 5:163
- Roberts JJ (1979) Antitumor platinum compounds. In: Antibiotics, V/2. Mechanism of action of antieukaryotic and antiviral compounds. Springer, Berlin Heidelberg, p 20

- Roberts JJ, Fraval NA (1980) Repair of cis-platinum(II)diamminedichloride-induced DNA damage and cell sensitivity. In: Cisplatin, current status and new developments. Academic Press, New York, p 57
- Roberts JJ, Pascoe JM (1972) Cross-linking of complementary strands of DNA in mammalian cells by antitumor platinum compounds. Nature 235: 282
- 71. Rosenberg B (1978) Platinum complex-DNA interactions and anticancer activity. Biochimie 60: 859
- Rosenberg B, Renshaw E, VanCamp L, Hartwick J, Drobník J (1967) Platinum-induced filamentous growth in *Escherichia coli*. J Bacteriol 93:716
- Salles B, Lesca C (1982) Induction of recA protein in *Escherichia coli* by three platinum(II) compounds. Biochem Biophys Res Commun 105: 202
- 74. Scovell WM, Kroos LR (1982) cis- and trans-Diamminedichloroplatinum(II) binding produces different structural changes on SV40 DNA. Biochem Biophys Res Commun 104: 1597
- 75. Srivastava RC, Froehlich J, Eichhorn GL (1978) The effect of platinum binding on the structure of DNA and its function in RNA synthesis. Biochimie 60:879
- 76. Szumiel I, Nias AHN (1976) Action of a platinum complex [cis-dichlorobis(cyclopentylamine)-platinum(II)] on Chinese hamster ovary cells in vitro. Chem Biol Interact 14:217
- Taylor DM, Jones JD, Robins AB (1973) Metabolism of platinum (14C)ethylendiamine dichloride in the rat. Biochem Pharmacol 22: 833
- 78. Tchernayev II, Krasnovskaya NN (1958) On the geometric isomers of dihydroxodiammine platinum(IV) dichlorides. Zh Neorg Khim 1958: 2024
- Teo B-K, Eisenberger P, Reed J, Barton JK, Lippard SJ (1978)
 Study of the binding of cis- and trans-dichlorodiammineplatinum(II) to calf thymus DNA by extended X-ray absorption fine structure spectroscopy. J Am Chem Soc 100: 3225
- 80. Thomson AJ (1974) The interactions of platinum compounds with biological molecules. In: Recent Results Cancer Res 48:38
- 81. Thomson AJ, Williams RJP, Reslová S (1972) The chemistry of complexes related to *cis*-Pt(NH₃)₂Cl₂, an anti-tumour drug. Structure and Bonding 11:2
- 82. Vonka V, Kutinová L, Drobník J, Bräuerová J (1972) Increase of Epstein-Barr-virus-positive cells in EB3 cultures after treatment with *cis*-dichlorodiammine-platinum(II). J Natl Cancer Inst 48:1277
- 83. Wiencke JK, Cervenka J, Paulus H (1979) Mutagenic activity of anticancer agent *cis*-dichlorodiammine platinum(II). Mutat Res 68:69
- 84. Zwelling LA, Kohn KW (1980) Effects of cisplatin on DNA and the possible relationships to cytotoxicity and mutagenicity in mammalian cells. In: Cisplatin, current status and new developments. Academic Press, New York, p 21
- 85. Zwelling LA, Anderson T, Kohn KW (1979) DNA-protein a and DNA interstrand cross-linking by *cis* and *trans*-platinum(II)diamminedichloride in L1210 mouse leukemia cells and relation to cytotoxicity. Cancer Res 39:365
- 86. Zwelling LA, Michaels S, Schwartz H, Dobson PP, Kohn KW (1981) DNA cross-linking as an indicator of sensitivity and resistance of mouse L1210 leukemia to cis-diamminedichloroplatinum(II) and L-phenylalanine mustard. Cancer Res 41:640